



For life science research use only. Not for use in diagnostic procedures. For *in vitro* use only.



LightMix[®] Modular *Escherichia coli* uidA

500

Cat.-No. 50-0641-96

Roche SAP n° 07 413 670 001

Kit with reagents for 96 PCR reactions 20 µl for detection of *E. coli* [lyophilized]

1. Content, Storage and Expiry

Storage at Arrival:

- 1 Vial orange cap 96 reactions *Escherichia coli* (lyophilized)
 - 1 Vial black cap Positive Control (32,000 copies, lyophilized)
- Store cooled or at ambient temperature
Do **not** freeze the lyophilized reagents.

- Lyophilized kits are stable for at least 6 months (4°C to 25°C in the dark). See lot-specific expiry date.
- Dissolved reagents are stable for at least 2 weeks if stored protected from light and cooled (4°C).
- Dissolved reagents can be stored long-term at -20°C (within expiry). Avoid multiple freeze-thaw cycles.
- Dissolved positive controls must be stored at -20°C. Avoid multiple freeze-thaw cycles.

2. Additional Reagents required

LightCycler[®] Multiplex DNA Master
or Roche LightCycler[®] 480 Probes Master (no instructions included)

Cat.-No. 07 339 585 001
Cat.-No. 04 707 494 001

3. Introduction

Bacterial meningitis in adults is caused mainly by *Neisseria meningitidis* or *Streptococcus pneumoniae*. Children might have *Haemophilus influenzae* while newborn can be also affected by *Streptococcus agalactiae* and *Escherichia coli*. *Listeria monocytogenes* is known to cause meningitis in elderly persons and newborns.

4. Description

A 87 bp long fragment from the beta-glucuronidase (*uidA*) gene of *Escherichia coli* is amplified with specific primers and detected with a Cyan500 labeled hydrolysis probe.

Note : The *uidA* gene is contained also in *Shigella*. *E.coli* and *Shigella* can be differentiated by testing for the *tnaA* and the *agaB* gene, which are missing in most *Shigella* strains.

5. Specification

This assay detects 10 target copies or less per reaction (plasmid DNA dilution).

6. Sample Material and Extraction

Typical sample type is cerebrospinal fluid (CSF). See ModularDx Document **Extraction Protocols**.

7. Material Safety Data (MSDS)

According to OSHA 29CFR1910.1200, Australia [NOHSC:1005, 1008 (1999)] and the EU Directives 67/548/EC and 1999/45/EC any products which do not contain more than 1% of a component classified as hazardous or classified as carcinogenic do not require a Material Safety Data Sheet (MSDS).

Product is not hazardous, not toxic, not IATA-restricted. Product is not from human, animal or plant origin. Product contains synthetic oligonucleotide primers and probes.



8. Instructions for Use

- Instrument programming see document
- Color Compensation see instructions in
- Pipetting instructions multiplex PCR see

ModularDx Programming

40-0320 Universal Color Compensation Hexaplex

ModularDx Multiplex

8.1. Programming Roche 480 Instruments

See the Instrument operator's manual for details. Start programming before preparing the solutions. The protocol consists of three program steps:

- 1: Denaturation: sample denaturation and enzyme activation
- 2: Cycling: PCR-amplification
- 3: Cooling: cooling the instrument

Detection Format 500 Channel

LightCycler® 480 Instrument:

LightCycler® 480 II Instrument:

cobas z 480 Analyzer (open channel):

Set Quant Factor 10 (default setting is 1)

450-500

440-488

no filter combination for Cyan500

| Program Step: | RT Step* | Denaturation | Cycling | | | Cooling |
|-----------------------------|----------|--------------|---------------------|----------|----------|----------|
| Parameter | | | | | | |
| Analysis Mode | None | None | Quantification mode | | | None |
| Cycles | 1 | 1 | 45 | | | 1 |
| Target [°C] | 55 | 95 | 95 | 60 | 72 | 40 |
| Hold [hh:mm:ss] | 00:05:00 | 00:05:00 | 00:00:05 | 00:00:15 | 00:00:15 | 00:00:30 |
| Ramp Rate [°C/s] 96 | 4.4 | 4.4 | 4.4 | 2.2 | 4.4 | 1.5 |
| Ramp Rate [°C/s] 384 | 4.6 | 4.6 | 4.6 | 2.4 | 4.6 | 2.0 |
| Acquisition Mode | None | None | None | Single | None | None |

* optional to combine with 1-Step RT-PCR

Table 1

8.2. Experimental Protocol

- **Sample material:** Use aqueous nucleic acid preparations (e.g. 'High Pure PCR Template Preparation Kit').
- **Negative control:** Always run at least one no-template control (NTC) - replace the template NA with water.
- **Positive control:** Run a positive control - replace the template NA with the provided Positive Control.

For an increased sensitivity use 10 µl sample per 20 µl reaction, in case that inhibition is likely to occur, e.g. extracts obtained from fecal samples, use 5 µl. For 10 µl reactions in 384 well plates use 5 µl /2.5 µl.

8.2.1. Preparation of Parameter-Specific Reagents (PSR, 96 reactions):

One reagent vial with an **orange** cap contains all primers and probe to run 96+ LightCycler® reactions.

Add 50 µl PCR-grade water to each reagent vial, mix the solution (vortex) and spin down.

For robotic pipetting the volume can be extended to 55 µl (signals will decrease by 10-20%).

► **Use 0.5 µl** reagent for a 20 µl PCR reaction.

8.2.2. Preparation of the Positive Control

Add 160 µl RNase/DNase-free Tris buffer or water to the vial with the **black** cap, for 10 µl sample add **320 µl**. Mix the solution by pipetting up and down 10 times. If vortexing spin down to collect the solution.

Notes: Opening of this vial may cause contaminations of the work-space (aerosol). Use of Tris buffer pH 8.0-8.5 increases the long-term stability in solution. Store dissolved controls frozen.

► **Use 5 µl** positive control (≈ 1,000 copies) for a 20 µl PCR reaction (or 10 µl if using 10 µl sample).

8.2.3. Preparation of the Reaction Mix

Multiply volumes by the number of reactions plus controls and one reserve and prepare in a cooled tube:

| For use with the Roche LightCycler® Multiplex DNA Master | | |
|--|--|----------------|
| for 5 µl extract | Component | 10 µl extract |
| 10.5 µl | Water , PCR-grade (colorless cap, provided with the Roche Master kit) | 5.5 µl |
| 0.5 µl | Reagent mix (parameter specific reagents containing primers and probes) | 0.5 µl |
| -- | Control Reaction and additional assays (Multiplex PCR) | -- |
| 4.0 µl | Roche Master (see Roche manual) | 4.0 µl |
| 15.0 µl | Volume of Reaction Mix | 10.0 µl |

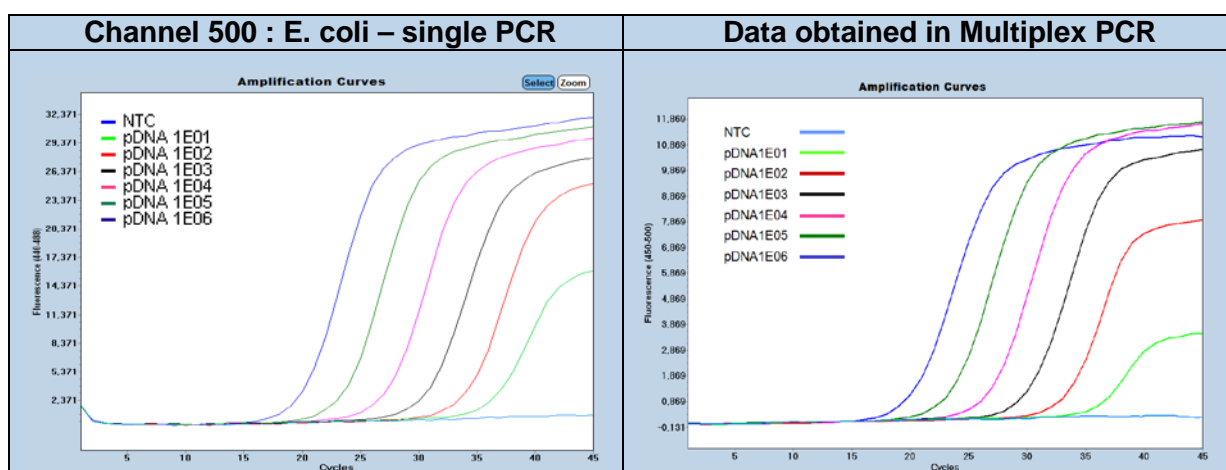
Table 2

Mix gently, spin down and **transfer 15 µl (10 µl)** per well.

Add 5 µl (10 µl) of sample or control to each well for a final reaction volume of 20 µl. Seal plate and centrifuge.

Start run

9. Typical Results (Data from LightCycler® 480 II system)



Dilution row 1E6 to 10 copies / reaction

Figure 1

10. Reading the Results

Perform data analysis as described in the operator's manual. For multiplex assays select the color compensation. We recommend using the Second Derivative Maximum method (Automated (F'' max)). View results in the 500 channel. The negative control (NTC) must show no signal.

| Channel 500 (sample) | Channel 660 Control Reaction | Channel 500 NTC Control | Result |
|------------------------|------------------------------|-------------------------|----------------------|
| No amplification | Detectable | Negative | Not detectable |
| Amplification Cp < 37* | Not relevant | Negative | E.coli. Positive |
| No amplification | Not detectable | Not relevant | PCR failure Repeat |
| Amplification signal | Not relevant | Positive | Contamination Repeat |

Note: cobas z 480 Analyzer signal levels are about 50% compared to LightCycler® 480 II results.
+ Recommendation : Define the cut-off 2-4 cycles higher than observed for 10 copies

Positive controls from for other assays may result in late positive signals (*E.coli* DNA background)

11. References

None

12. Multiplex PCR Compatibility (Meningitis Panel or Shigella testing)


This *Escherichia coli* kit can be combined with other assays up to 6plex reactions including an internal control (IC) or a spiked extraction control (for example PhHV) as depicted below:

| Meningitis Multiplex PCR and Instrument Compatibility | | | | | | 480 II | z 480 | LC96 | LC2.0 | Nano |
|---|--------|----------|-------|------------|------|--------|-------|------|-------|------|
| Color Compensation 40-0320 is mandatory for Multiplex PCR | | | | | | | | | | |
| 500 | 530 | 580 | 610 | 640 | 660 | | | | | |
| uidA | L.mono | S.pneumo | N.men | H.influenz | PhHV | X | | | | |
| uidA | L.mono | S.pneumo | S.aga | H.influenz | PhHV | X | | | | |
| uidA | tnaA | Shigella | agaB | | PhHV | X | | | | |

Table 3

13. Version History

| | | |
|---------|--|------------|
| V140404 | Release version | 2014-04-14 |
| V140909 | Editorial changes | 2014-09-09 |
| V151001 | 2015 protocol Multiplex Master, 5/10 µl extract and 60°C acquisition | 2015-10-01 |
| V161001 | False positive results with other plasmid based controls possible | 2016-01-07 |
| V160313 | 1. Storage of controls, 8.2.2 buffer, 8.2.3 wording | 2016-06-10 |

| Certificate of Analysis (CoA) | | | | | | |  |
|--|--------------|------------|------------|-----------|------------|------------|---|
| Lot n° | | | | | | | |
| Expiry : | | | | | | | |
| Dilution | 1E6 | 1E5 | 1E4 | PC | 1E2 | 1E1 | passed |
| Cp range | 19-22 | 22-25 | 25-28 | 28-32 | 32-35 | 35-38 | |
| Measured Signal level | 31 | | | | | | |
| Measured | 80-120 | | | | | | |
| Negatives | 10/10 | | | | | | ✓ |
| Note: Cp (crossing point) values collected with pDNA (single target PCR). Fluorescence (FL) levels depend on instrument settings and may vary. The Cp values will vary from instrument to instrument by up to 2 cycles, while the distance between two dilution steps should be relative constant (Δ Cp). | | | | | | | |
| QC Acceptance Date: | | | | YYYYMMDD | | | |
| We, the undersigned, certify that the product designated above has been obtained in accordance with the rules of production and quality control. | | | | | | | |
| Name(s) : | | | | | | | |

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