

LightMix[®] Kit *human Cytomegalovirus (hCMV) EC*

Cat.-No. 40-0562-32

Internal Control (IC) changed to Extraction Control (EC), 32 rxns/vial

Kit with reagents for detection of *human Cytomegalovirus* genomic DNA using the Roche Diagnostics LightCycler[®] 1.x / 2.0 / 480 II Instruments or cobas z 480 Analyzer.

Lyophilized mix of primers and probes for a total of 96 reactions with a final volume of 20 µl each. **Store protected from light at room temperature (18-25°C), do NOT freeze!**

Instructions for use with the LightCycler[®] 1.x / 2.0 Instruments see pages 4-5

Instructions for use with the LightCycler[®] 480 II and cobas z 480 Analyzer see pages 6-7

1. Introduction

Human Cytomegalovirus (CMV), also known as Human Herpesvirus 5 (HHV-5), is a double stranded DNA virus and member of the herpes virus family. Primary infections of immunocompetent hosts are normally without symptoms; some individuals develop fever, and a very small percentage suffers from a mononucleosis syndrome or a mild hepatitis. About 50-80% of the population is positive for hCMV. Infections of unborn and young children as well as of immuno compromised individuals may cause severe symptoms and can be life threatening.

CMV will be tested in case of signs of mononucleosis and absence of EBV or symptoms of hepatitis and negative results for HAV, HBV or HCV. Diagnosis of hCMV infections is based on serology, virus culture or PCR. Preferred target genes for the Real-Time-PCR are the immediate early (MIE)¹, the polymerase (UL54), glycoprotein B (UL55)², the phosphoprotein pp65 (UL83)³ genes or intron UL93⁴ for which several TaqMan and LightCycler[®] assays have been published¹⁻⁴.

This LightMix[®] kit provides a fast, easy and accurate system to identify this target in a nucleic acid extract. The internal control (IC) has been changed to a spiked extraction control (sEC) to monitor a successful extraction and demonstrate the ability to run an amplification reaction (thus absence of PCR inhibition). The kit has been tested with 'LightCycler[®] FastStart DNA Master HybProbe' only.

We recommend to use the 'Extraction Control' procedure; for compatibility with the former procedure the use as IC is also described. Target and control primer/probe sequences remained unchanged.

¹ Nitsche A, Steuer N, Schmidt CA, Landt O, Siebert W. Different real-time PCR formats compared for the quantitative detection of human cytomegalovirus DNA. Clin Chem. 45 (1999) 1932-1937

² Detection of cytomegalovirus DNA in human specimens by LightCycler PCR. Schaade L, Kockelkorn P, Ritter K, Kleines M. J Clin Microbiol. 38 (2000) 4006-4009

³ Detection of HSV-1/-2, VZV, EBV and CMV by a set of LightCycler PCRs Performed in Parallel within a Single Run. Stöcher, M., Leb, V. Bosic, M., Kessler, H. Halwachs-Baumann, G., Landt, O., Stekel, H. Berg, J., J Clin Virol 26 (2003) 85-93

⁴ Schalasta G, Eggers M, Schmid M, Enders G. Analysis of human cytomegalovirus DNA in urines of newborns and infants by means of a new ultrarapid real-time PCR-system. J Clin Virol. 2000 Oct 1;19(3):175-185

2. Description

A 226 bp long fragment from the *hCMV* Glycoprotein B gene (UL55) is amplified with specific primers. The resulting PCR fragment is detected with LightCycler[®] Red 640 labeled hybridization probes. The sequences used in this device are derived from the assay as published by Schaade et al.³

The control reaction generates an additional product of 278 bases which is detected with LightCycler[®] Red 690 labeled hybridization probes. This second PCR has no visible impact on the hCMV specific reaction and will even fail in the presence of higher amounts of hCMV target (1,000 copies and more).

The use of a color compensation file generated with the ColorCompensation kit 40-0318 is a prerequisite to run the duplex reaction *hCMV* and the control.

The supplied standard row allows to determine the linear range of the reaction and to estimate the quantity of the target sequence in unknown samples.

3. Set Contents

- 3 Vials with **blue** cap containing premixed lyophilized primers and probes for 32 reactions hCMV
- 1 Standard row with 6 lyophilized standards of hCMV from 10^1 to 10^6 target equivalents per rxn
- 1 Sealing foil for the standard row
- 3 Vials with **white** cap containing premixed lyophilized primers and probes for 32 reactions EC
- 1 Vial with **white** cap containing the Extraction Control Target (ECT) with 2×10^6 copies (total)
- 1 Vial with **black** cap containing the stabilizer for the No Template Control (NTC)
- 1 Certificate of Analysis (CoA) with lot-specific data

4. Additional Reagents and items required

	Roche Diagnostics
Color Compensation HybProbe order n°40-0318-00	Cat.-No. 05 997 704 001
LightCycler® FastStart DNA Master HybProbe	Cat.-No. 03 003 248 001
LightCycler® Capillaries (20 µl) (LightCycler® 1.x / 2.0 Instruments)	Cat.-No. 04 929 292 001
LightCycler® 480 Multiwell Plate 384, white (LightCycler® 480 Instrument)	Cat.-No. 04 729 749 001
or LightCycler® 480 Multiwell Plate 96, white (LightCycler® 480 Instrument)	Cat.-No. 04 729 692 001

For use in LightCycler® 1.x Instruments with software version 3.5.3 read channel F2 instead of 640 and F3 instead of channel 705. We recommend to upgrade to software version 4.10 or higher.

4.1. Optional Additional Reagents

High Pure Viral Nucleic Acid Kit	Cat.-No. 11 858 874 001
or High Pure PCR Template Preparation Kit	Cat.-No. 11 796 828 001

5. Product Characteristics

PCR results (activation, 50 cycles and melting curve) are obtained within 50 minutes with the capillary based LightCycler® 1.x / 2.0 Instruments and within 80 minutes with '480' plate based Instruments.

Sensitivity

These reagents detect 10 copies of hCMV DNA using the Roche 'LightCycler® FastStart DNA Master HybProbe' (in an exemplary system, using cloned targets as reference).

Measuring range

The linear measuring range of the assay is 10^2 to 10^6 copies of hCMV DNA using the Roche 'LightCycler® FastStart DNA Master HybProbe'.

Storage and Stability

- Lyophilized reagents are stable for at least 6 months after shipment when stored protected from light at room temperature (18-25°C). See expiry date on the product label.
- **Do not freeze** lyophilized reagents.
- Dissolved reagents are stable for at least 10 days when stored protected from light and refrigerated (4°C).

6. Experimental Protocol

Start programming before preparing the solutions. See the Instrument operator's manual for details.

6.1. Preparation of parameter-specific reagents and reagents for the EC (32 reactions):

One reagent vial with a **blue** cap contains primers and probes to run 32 reactions hCMV.
One reagent vial with a **white** cap contains primers and probes to run 32 reactions control reaction.

Add **66 µl** PCR-grade water to each reagent vial, mix the solution (vortex) and spin down.

► Use **2 µl** reagent for a 20 µl PCR reaction.

This solution is stable at least ten days when stored refrigerated at 4°C. Avoid prolonged exposure to light.

6.2. Preparation of Extraction Control Target (ECT):

Add **1,200 µl** PCR-grade water to the vial with **white** cap containing the Extraction Control Target

6.3. Sample Preparation:

Before extraction add **10 µl** of ECT (vial **white** cap) to the sample. **Skip if IC procedure is used.**
Perform nucleic acid preparation as described in the protocol of the extraction kit used (see 4.1).

6.4. Preparation of No Template Extraction (NTC):

Add **900 µl** PCR-grade water to the **NTC** (vial **black** cap) and add **100 µl** of ECT (vial **white** cap).

► Use **5 µl** No Template Control DNA for a 20 µl PCR reaction.

6.5. Preparation of the standard row

The target DNA is provided in 6 different quantities to yield from 10 to 10^6 target molecules in 5 µl once resolved. Start with the lowest concentrated standard (first tube from the extended lip). Use the pipette tip to punch a hole through the sealing foil. Add **40 µl** of **NTC** (vial **black** cap) to each well. **Use water if the IC procedure is selected.** Mix the target DNA by pipetting the solution up and down 10 times.



► Use **5 µl** standard for a 20 µl PCR reaction.

This standard solution is not long-term stable. Use only fresh prepared solutions only. After adding the target DNA to the reaction mix use the provided sealing foil to close the vials in order to avoid contaminations.

6.6. Preparation of the Reaction Mix

Include at least one 'No Template Control' (NTC) and one Positive Control. In a cooled reaction tube, prepare the reaction mix by multiplying each volume for a single reaction by the number of reactions (samples plus controls) to be cycled, plus one additional reaction.

sEC Procedure	For use with the Roche FastStart Master	IC Procedure
Single reaction	Component	Single reaction
6.4 µl	Water, PCR-grade (colorless cap, provided with the Roche Master kit)	5.9 µl
2.6 µl	Mg ²⁺ solution 25 mM (blue cap, provided with the Roche FastStart kit)	2.6 µl
2.0 µl	Reagent mix (parameter specific reagents, primers and probes, see 6.1.)	2.0 µl
2.0 µl	EC mix (EC reagents containing primers, probes, see 6.1.)	2.0 µl
---- µl	ECT Control Target (vials white cap)	0.5 µl
2.0 µl	Roche Master (red cap, for preparation see Roche manual)	2.0 µl
15.0 µl	Volume of reaction mix	15.0 µl

Table 1

To run the assay without the control reaction substitute ECT with 0.5 µl PCR-grade water.

Mix gently, spin down and **transfer 15 µl** of the reaction mix to a capillary or well.

Add 5 µl of sample or standard to each capillary or well for a final reaction volume of 20 µl.

Close the capillaries / attach a foil to the multiwell plate and seal, and spin down.

Start run.

7. LightCycler® 1.x / 2.0 Instruments

7.1. Programming

The protocol consists of four program steps

- 1: Denaturation: sample denaturation and enzyme activation
- 2: Cycling: PCR-amplification of the target DNA
- 3: Melting: melting curve analysis for identification of the PCR product derived from the target DNA
- 4: Cooling: cooling the instrument

Program Step:	Denaturation	Cycling			Melting			Cooling
Parameter								
Analysis Mode	None	Quantification mode			Melting Curves mode			None
Cycles	1	50			1			1
Target [°C]	95	95	62	72	95	40	85	40
Hold [hh:mm:ss]	00:10:00	00:00:05	00:00:05	00:00:15	00:00:20	00:00:20	00:00:00	00:00:30
Ramp Rate [°C/s]	20	20	20	20	20	20	0.2	20
Sec Target [°C]	-	-	55	-	-	-	-	-
Step Size [°C]	-	-	0.5	-	-	-	-	-
Step Delay (Cycles)	-	-	1	-	-	-	-	-
Acquisition Mode	None	None	Single	None	None	None	Continuous	None

(Melting not relevant for detection)

Table 2

7.2. Data Analysis

Switch the color compensation mode on. If this mode is not enabled run the color compensation program. Follow the instructions in the manual of the 'LightMix® Kit – Color Compensation HybProbes'.

We recommend using the Second Derivative Maximum method (Automated (F" max)). The cycle number of the Crossing Point (Cp) of each sample is calculated automatically. The Fit Points method is more-error prone due to the user's influence.

View hCMV data in channel 640, Quantification mode. The Negative Control must show no signal.

If the control reaction is used, view data in channel 705, Quantification mode. The negative control and the low-concentrated hCMV DNA samples (10 to 1,000 copies) should show an amplification curve for the IC/EC with a Cp at approximately cycle 30.

The provided standard row with 10⁶ copies/rxn to 10 copies/rxn of hCMV should have Cp values between cycles 18 and 35 (Cp values calculated with Second Derivative Maximum method).

For use in LightCycler® 1.x Instruments use channel F2 instead of channel 640 and channel F3 instead of channel 705 for detection.

7.3. Sample Data – Typical Results

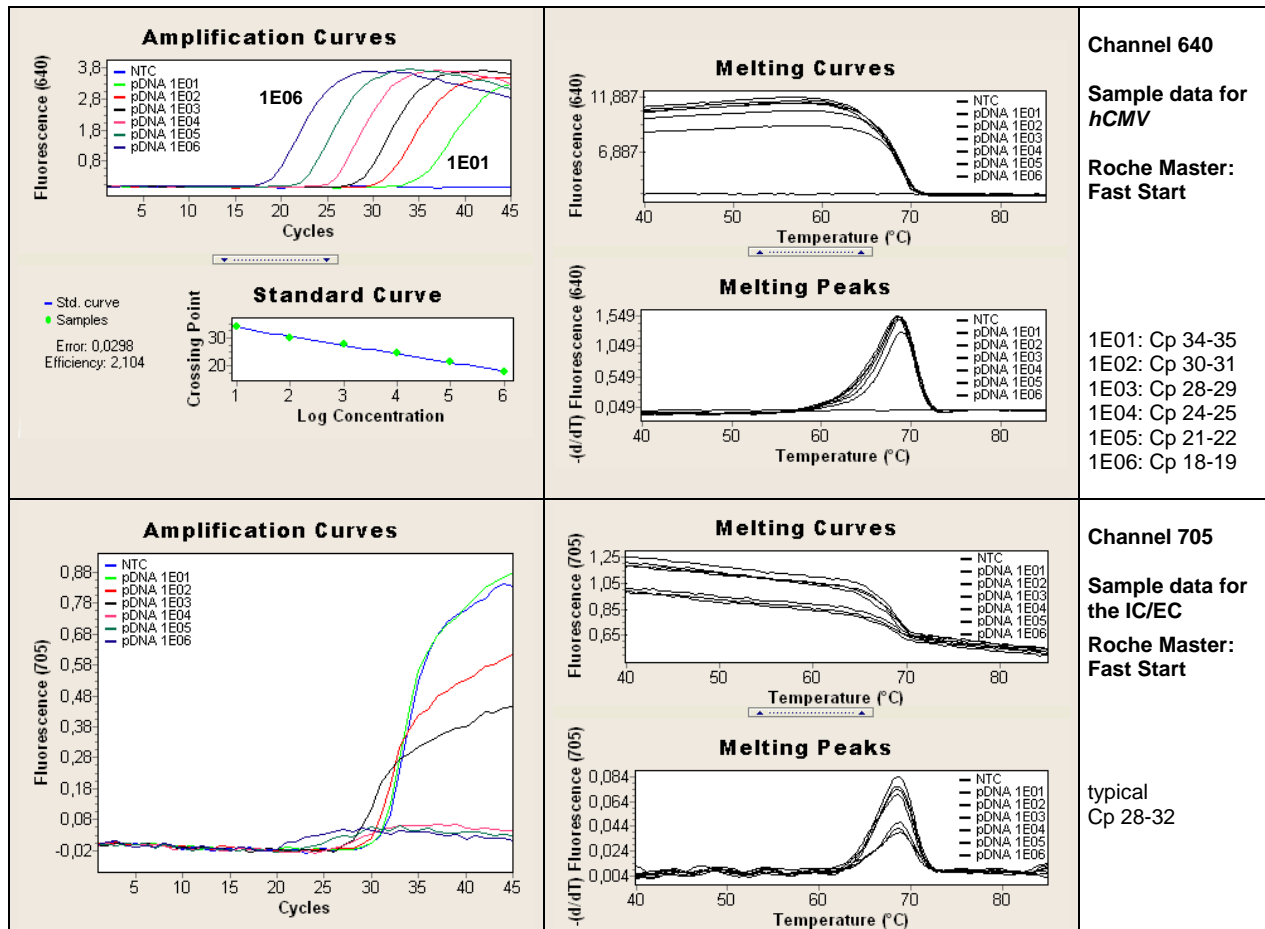


Fig.1. LightCycler® 2.0 sample data for the *hCMV* detection system.

Upper panels: Left panel channel 640 quantification mode (Second Derivative Maximum) with the standard row *hCMV*. Right panel channel 640 melting analysis for *hCMV* (not relevant for detection).

Lower panels: Left panel channel 705 quantification mode (Second Derivative Maximum) for the control reaction. Right panel channel 705 melting analysis for the control reaction (not relevant for detection).

7.4. Interpretation of Data

Sample 640 <i>hCMV</i>	Sample 660 Control	Channel 640 Positive Control	Channel 640 Negative Control	Result (warnings)
No amplification	Cp 28-31	amplification	negative	Negative (not detectable)
Cp < 37*	not relevant	amplification	negative	Positive for <i>hCMV</i>
no amplification	not detectable	amplification	not relevant	PCR failure, repeat experiment
not relevant	not relevant	no amplification	not relevant	PCR failure, repeat experiment
not relevant	not relevant	not relevant	positive	Contamination, repeat experiment

Table 3 Typical analysis results with LightCycler® 2.0 Instrument

* The cut off value is a recommendation only - this value has to be defined by the user. Compare with the lot specific values for 10 copies as reported in the Certificate of Analysis (CoA). 1-2 cycles later than observed for 10 copies corresponds to ~5 copies.

8. LightCycler® 480 II Instrument and cobas z 480 Analyzer

Using the kit with the LC 480 (Version I) will yield no or very weak signals for the control reaction while the signals in channel 640 for *hCMV* are not affected.

8.1. Programming

The protocol consists of four program steps

- 1: Denaturation: sample denaturation and enzyme activation
- 2: Cycling: PCR-amplification of the target DNA
- 3: Melting: melting curve analysis for identification of the PCR product derived from the target DNA
- 4: Cooling: cooling the instrument

Detection Format:

LightCycler® 480 II Instrument: 465-510, 498-640, 498-660

cobas z Analyzer: 465-510, 498-645, 498-700

Program Step:	Denaturation	Cycling			Melting			Cooling
Parameter								
Analysis Mode	None	Quantification mode			Melting Curves mode			None
Cycles	1	50			1			1
Target [°C]	95	95	62	72	95	40	85	40
Hold [hh:mm:ss]	00:10:00	00:00:05	00:00:05	00:00:15	00:00:30	00:01:00	00:00:00	00:00:30
Ramp Rate [°C/s] 96	4.4	4.4	2.2	4.4	4.4	1.5	-	1.5
Ramp Rate [°C/s] 384	4.6	4.6	2.4	4.6	4.6	2.0	-	2.0
Sec Target [°C]	-	-	55	-	-	-	-	-
Step Size [°C]	-	-	0.5	-	-	-	-	-
Step Delay (Cycles)	-	-	1	-	-	-	-	-
Acquisition Mode	None	None	Single	None	None	None	Continuous	None
Acquisitions [per °C]	-	-	-	-	-	-	1	-

(Melting not relevant for detection)

Table 4

8.2. Data Analysis

Note: cobas z 480 Instruments signal levels are about 50% compared to LightCycler® 480 II results.

Switch the color compensation mode on. If this mode is not enabled run the color compensation program. Follow the instructions in the manual of TIB ColorCompensation HybProbe.

Perform data analysis, as described in the LightCycler® Instrument operator's manual.

We recommend using the Second Derivative Maximum method (Automated (F'' max)). The cycle number of the Crossing Point (Cp) of each sample is calculated automatically. The Fit Points method is more-error prone due to the user's influence.

View *hCMV* data with Filter Combination 498-640 (498-645), Quantification mode. The negative control (NTC) must show no signal.

If the control reaction is used, view data in Filter Combination 498-660 (498-700), Quantification mode. The negative control and the low-concentrated *hCMV* DNA samples (10 to 1,000 copies) should show an amplification curve for the IC/EC with a Cp at approximately cycle 30.

The provided standard row with 10⁶ copies/rxn to 10 copies/rxn of *hCMV* should have Cp values between cycles 18 and 35 (Cp values calculated with Second Derivative Maximum method).

8.3. Sample Data – Typical Results

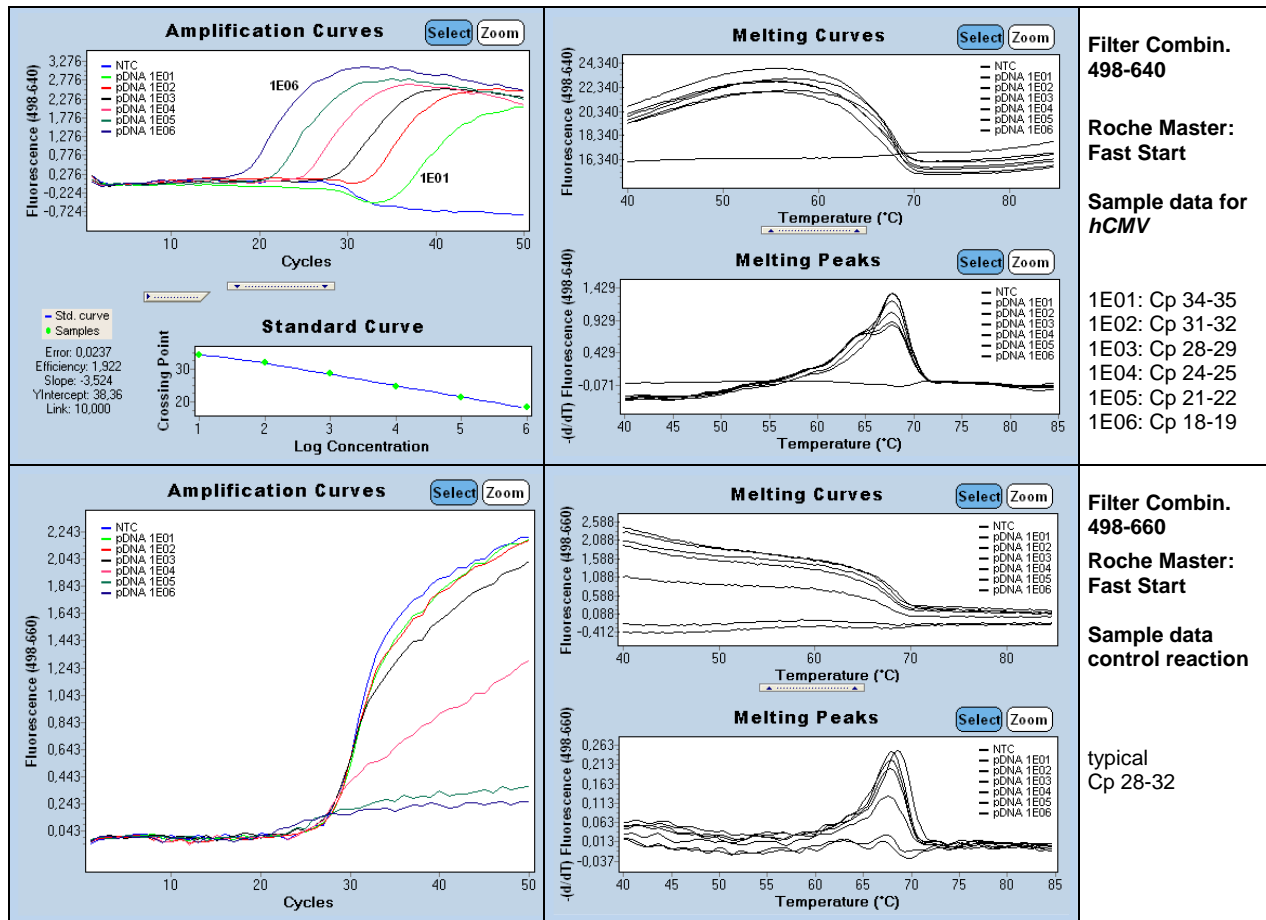


Fig.2. LightCycler® 480 II sample data for the hCMV detection system.

Upper panels: Left panel filter combination 498-640 quantification mode (Second Derivative Maximum) obtained with the standard row for hCMV. Right panel filter combination 498-640 melting analysis for hCMV (not relevant for detection).
Lower panels: Left panel filter combination 498-660 quantification mode (Second Derivative Maximum) for the control reaction. Right panel channel filter combination 498-660 melting analysis for the control reaction (not relevant for detection).

8.4. Interpretation of Data

Sample 640 hCMV	Sample 660 Control	Channel 640 Positive Control	Channel 640 Negative Control	Result (warnings)
No amplification	Cp 28-31	amplification	negative	Negative (not detectable)
Cp < 37*	not relevant	amplification	negative	Positive for hCMV
no amplification	not detectable	amplification	not relevant	PCR failure , repeat experiment
not relevant	not relevant	no amplification	not relevant	PCR failure , repeat experiment
not relevant	not relevant	not relevant	positive	Contamination , repeat experiment

Table 5 Typical analysis results with LightCycler® 480 II Instrument

* The cut off value is a recommendation only - this value has to be defined by the user. Compare with the lot specific values for 10 copies as reported in the Certificate of Analysis (CoA). 1-2 cycles later than observed for 10 copies corresponds to ~5 copies.

9. Conversion Factor

The amount of virus per sample (Viral Load) is usually reported in copies / ml while PCR reports copies per reaction. The conversion factor between both depends on sample and extraction volumes. Extraction starts usually from less than one milliliter and PCR does not use the total extracted volume.

The viral load (VL) can be calculated using the following general formula:

$$VL \text{ [copies/ml]} = MV \times EVF \times SF$$

where:

VL	=	Viral Load
MV	=	Measured Value [copy number per reaction]
EVF	=	Extraction Volume Factor [Final extraction volume / PCR sample volume]
SF	=	Sample Factor [1,000 µl / extracted volume of clinical sample]

For example, extracting from 200µl of clinical sample results in a correction factor of 5. Using 5 µl extract from a total elution volume of 100 µl results in a correction factor of x 20, resulting in a conversion factor of 100.

$$VL \text{ [copies/ml]} = \text{Measured Value [copy number per reaction]} \times 100$$

10. Material Safety Data

According to OSHA 29CFR1910.1200, Commonwealth of Australia [NOHSC:1005, 1008 (1999)] and the European Union Directives 67/548/EC and 1999/45/EC any products which not contain more than 1% of a component classified as hazardous or classified as carcinogenic do not require a Material Safety Data Sheet (MSDS).

Product is not hazardous, not toxic, not IATA-restricted. Product is not from human, animal or plant origin. Product contains synthetic oligonucleotide primers and probes.

11. Version History

Notes in red mark events require to change procedures

V110901
V121024

Increased MgCl₂ concentration to 4.25mM

Chapter 9. Conversion Factor included.
Instrument cobas z 480 analyzer included.
Roche Color Compensation kit removed

V130628
V140414

MSDS included, editorial changes

Change from 16 reactions / vial to 32 reactions / vial
Change Internal Control (IC) to Extraction Control (EC)
480 instruments number of acquisitions corrected to 1

Roche SAP order n° 05943566001

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These reagents were developed and manufactured by TIB MOLBIOL GmbH, Berlin, Germany.
LightCycler® hybridization probes produced under license from Roche Diagnostics GmbH.

